

This Page Is Inserted by IFW Operations
and is not a part of the Official Record

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

**As rescanning documents *will not* correct images,
please do not report the images to the
Image Problem Mailbox.**

INTER LIBRARY LOAN REQUEST FORM

Borrower's Name: SHARON FOLEY Org. or A.U. 1648 Phone 308-3983
 Serial Number 09/506011 Date of Request 4/27/01 Date Needed By 5/5/01

Please Attach Copy Of Abstract, Citation, Or Bibliography, If Available. Please Provide Complete Citation. Only One Request Per Form.

Author/Editor:
Journal/Book Title:
Article Title: <u>SEE ATTACHED</u>
Volume (Issue):
Pages:
Year of Publication:
Publisher:
Remarks:
COMPLETED

STIC Use Only

Accession Number: 343,562

LIBRARY ACTION	LC		NAL		NIH		NLM		NBS		PTO		OTHER	
	1st	2nd	1st	2nd	1st	2nd	1st	2nd	1st	2nd	1st	2nd	1st	2nd
Local Attempts														
Date					<u>5/3</u>									
Initials					<u>SMP</u>									
Results					<u>NBS</u>									
Examiner Called														
Page Count														
Money Spent														

Provided By: Source and Date

Ordered From: Source and Date

Remarks/Comments

1st & 2nd denotes time taken to a library

O/N - Under NLM means Overnight Service

FX - Means Faxed to us

are Na sulphonated polystyrene or styrene-maleic acid copolymer.

USE - The cleaning and regeneration of body fluids (blood, plasma, serum, ascites, hydrothorax of patients with renal insufficiency or tumours by selective removal or beta 2 micro-globulin without taking out albumin or posing a need for replacement of body fluids. @

227 ANSWER 12 OF 12 CANCERLIT

ACCESSION NUMBER: 85609630 CANCERLIT

DOCUMENT NUMBER: 85609630

TITLE: RECEPTOR-MEDIATED ENDOCYTOSIS BY NORMAL AND PROLIFERATING HEPATOCYTES AND LIPOSOMAL DRUG DELIVERY.

AUTHOR: Wolkoff A W; Stockert R J; Schein P S

CORPORATE SOURCE: Liver Res. Center, Albert Einstein Coll. of Medicine, 1300 Morris Park, Bronx, New York, NY 10461.

SOURCE: Dev Oncol, (1984). Vol. 24, pp. 278-91.

DOCUMENT TYPE: Journal; Article; (JOURNAL ARTICLE)

FILE SEGMENT: ICDB

LANGUAGE: English

ENTRY MONTH: 198507

AB A specific liver cell membrane receptor, hepatic binding protein (HBP), is necessary for the uptake of asialoglycoproteins by hepatocytes. Isolated perfused rat liver pre-infused with anti-HBP IgG exhibited 80% reduction in asialoorosomucoid (ASOR) uptake, but no change in bilirubin uptake,

Searcher : Shears 308-4994

Nov/27

DEVELOPMENTS IN ONCOLOGY

- F.J. Cleton and J.W.I.M. Simons, eds., *Genetic Origins of Tumour Cells*
ISBN 90-247-2272-1
- J. Aisner and P. Chang, eds., *Cancer Treatment Research*
ISBN 90-247-2358-2
- B.W. Ongerboer de Visser, D.A. Bosch and W.M.H. van Woerkom-Eykenboom, eds.,
Neuro-oncology: Clinical and Experimental Aspects
ISBN 90-247-2421-X
- K. Hellmann, P. Hilgard and S. Eccles, eds., *Metastasis: Clinical and Experimental Aspects*
ISBN 90-247-2424-4
- H.F. Seigler, ed., *Clinical Management of Melanoma*
ISBN 90-247-2384-4
- P. Correa and W. Haenszel, eds., *Epidemiology of Cancer of the Digestive Tract*
ISBN 90-247-2601-8
- L.A. Liotta and I.R. Hart, eds., *Tumour Invasion and Metastasis*
ISBN 90-247-2611-5
- J. Bándóczy, ed., *Oral Leukoplakia*
ISBN 90-247-2655-7
- C. Tijsen, M. Halprin and L. Endtz, eds., *Familial Brain Tumours*
ISBN 90-247-2691-3
- F.M. Muggia, C.W. Young and S.K. Carter, eds., *Anthracycline Antibiotics in Cancer*
ISBN 90-247-2711-1
- B.W. Hancock, ed., *Assessment of Tumour Response*
ISBN 90-247-2712-X
- D.E. Peterson, ed., *Oral Complications of Cancer Chemotherapy*
ISBN 0-89838-563-6
- R. Mastrangelo, D.G. Poplack and R. Riccardi, eds., *Central Nervous System Leukemia. Prevention and Treatment*
ISBN 0-89838-570-9
- A. Pollack, ed., *Human Leukemias. Cytochemical and Ultrastructural Techniques in Diagnosis and Research*
ISBN 0-89838-585-7
- W. Davis, C. Maltoni and S. Tanneberger, eds., *The Control of Tumor Growth and its Biological Bases*
ISBN 0-89838-603-9
- A.P.M. Heintz, C.Th. Griffiths and J.B. Trimpos, eds., *Surgery in Gynecological Oncology*
ISBN 0-89838-604-7
- M.P. Hacker, E.B. Double and I. Krakoff, eds., *Platinum Coordination Complexes in Cancer Chemotherapy*
ISBN 0-89838-619-5
- M.J. van Zwieten, *The Rat as Animal Model in Breast Cancer Research: A Histopathological Study of Radiation- and Hormone-Induced Rat Mammary Tumors*
ISBN 0-89838-624-1
- B. Löwenberg and A. Hagenbeek, eds., *Minimal Residual Disease in Acute Leukemia*
ISBN 0-89838-630-6
- C.J.H. van de Velde and P.H. Sugarbaker, eds., *Liver Metastasis*
ISBN 0-89838-648-5

LIVER METASTASIS

Basic aspects, detection and management

edited by

Cornelis J.H. VAN DE VELDE, MD, PhD
*Department of Surgery, University Hospital
Leiden, The Netherlands*

Paul H. SUGARBAKER, MD
*Colorectal Cancer Section, National Institutes of Health
Bethesda, Maryland, USA*

1984 MARTINUS NIJHOFF PUBLISHERS
a member of the KLUWER ACADEMIC PUBLISHERS GROUP
BOSTON / DORDRECHT / LANCASTER



PREFACE

E. CADY

Hepatic metastases present one of the major therapeutic challenges of cancer patient management, for it is the destruction of vital organ function that makes cancer fatal, not local tumor growth. The process of tumor cell dislodgement from the primary cancer, their spread through the lymphatic and hematogenous channels, their lodgement in distant sites, and their subsequent progressive growth tax our comprehension and frustrate our therapies. The proceedings of this International Congress on Hepatic Metastasis address these aspects of metastases to the liver, and predominantly focus on metastatic colon cancer because of its frequency, its prominent hepatic only pattern of spread, and enticing preliminary data about prevention and control of small subsets of the afflicted population. Predictably, the "false technologies" of Dr. Lewis Thomas that involve surgical, radiotherapeutic and chemotherapeutic attack on these metastases after elaborate diagnostic studies take precedence because of the clinical imperatives of sick patients. This is displayed in the preponderance of papers and interest in various diagnostic scanning techniques by means of radioisotopes, radiographically useful dyes, biochemical markers, interest in developing accurate staging systems to categorize patients for therapeutic comparisons, and interest in elaborate, and expensive, technology to increase the effectiveness of chemotherapeutic agents that are of limited benefit with simple intravenous administration.

Behind this clinical enthusiasm, however, lies the research to develop the "true technology," in Thomas' words, that will prevent such clinical catastrophes as hepatic metastases. The first inkling of such a "true technology" in liver cancer is the recent development of hepatitis immunization to prevent subsequent hepatocellular carcinoma areas of the world. In hepatic metastases from colon cancer, several



Distributors

for the United States and Canada: Kluwer Academic Publishers, 190 Old Derby Street, Hingham, MA 02043, USA
for the UK and Ireland: Kluwer Academic Publishers, MTP Press Limited, Falcon House, Queen Square, Lancaster LA1 1RN, England
for all other countries: Kluwer Academic Publishers Group, Distribution Center, P.O. Box 322, 3300 AH Dordrecht, The Netherlands

Library of Congress Cataloging in Publication Data

Liver metastasis.

(Developments in oncology)

Proceedings of the International Congress on Hepatic Metastasis, sponsored by the University Hospital of Leiden and the National Cancer Institute, USA; and held in Leiden, May 24-26, 1984.

Includes bibliographies and index.

1. Liver--Cancer--Congresses. 2. Metastasis--Congresses. 3. Cancer invasiveness--Congresses.
- I. Velde, Cornelis J. H. van de. II. Sugarbaker, Paul H. III. International Congress on Hepatic Metastasis. (1984 : Leiden, Netherlands)
- IV. Academisch Ziekenhuis (Leiden, Netherlands)
- V. National Cancer Institute (U.S.) VI. Series.

RC280.L5L385 1984 616.99'436 84-16621
 ISBN 0-89838-684-5

ISBN 0-89838-684-5 (this volume)

Book Information

This publication is based upon a Boerhaave course organized by the Faculty of Medicine, University of Leiden, The Netherlands in co-operation with the National Cancer Institute, Bethesda, Maryland, USA

Copyright

© 1984 by Martinus Nijhoff Publishers, Dordrecht.

All rights reserved. No part of this publication may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, mechanical, photocopying, recording, or otherwise, without the prior written permission of the publishers,

Martinus Nijhoff Publishers, P.O. Box 163, 3300 AD Dordrecht, The Netherlands.

PRINTED IN THE NETHERLANDS

RECEPTOR-MEDIATED ENDOCYTOSIS BY NORMAL AND PROLIFERATING HEPATOCYTES AND LIPOSOMAL DRUG DELIVERY

ALLAN W. WOLKOFF, M.D., RICHARD J. STOCKERT, Ph.D. AND PHILIP S.
SCHEIN, M.D.

Receptor-mediated endocytosis is a process common to many species and cell types. One of the best characterized systems in which this process occurs is that of the hepatocyte receptor for asialoglycoproteins (1). This receptor was first described by Ashwell and Norell in studies of plasma disappearance of ceruloplasmin (2). In these studies performed in rats, they determined that native ceruloplasmin had a circulating half-life of 55 hours. Like virtually all mammalian plasma proteins, with the exception of albumin, ceruloplasmin is a glycoprotein consisting of a protein core with complex carbohydrate side-chains attached via aspartate residues. The terminal carbohydrate in these chains is sialic acid; the penultimate is galactose. Removal of sialic acid, exposing galactosyl residues, resulted in a reduction in circulating half-life to minutes rather than hours. Plasma clearance of asialoceruloplasmin as well as most other asialoglycoproteins represents uptake into hepatocytes. This uptake is mediated by a specific liver cell membrane receptor, hepatic binding protein (HBP) (3).

HBP is a membrane glycoprotein which has been solubilized in detergent and purified from rat, rabbit and human liver. As demonstrated in studies performed in isolated perfused rat liver, HBP is necessary for uptake of asialoglycoproteins by hepatocytes (4). In these studies, rat liver was first perfused with 100 mg of non-immune goat IgG (Figure 1). Following IgG infusion, a mixture of 125I-Asialoorosomucoid (ASOR), ³H-bilirubin and ¹³¹I-albumin was injected as a small bolus into the portal vein. Albumin was used as a non-transported reference. Its extracellular space of distribution is that of bilirubin, which circulates bound to it, and is similar to

that of ASOR, a protein of comparable molecular weight. Following injection, all effluent coming from the hepatic vein was collected in aliquots every 1-2 seconds without recirculation. In this way, uptake of bilirubin and ASOR during a single pass through the liver could be quantitated. Following this study, anti-HBP IgG was infused and the study repeated (Figure 2). Analysis revealed that uptake of ASOR was reduced by over 80% following anti-HBP infusion, while bilirubin uptake did not differ from control. These studies also revealed that uptake of bilirubin which occurs by facilitated diffusion rather than by endocytosis is independent of uptake of ASOR.

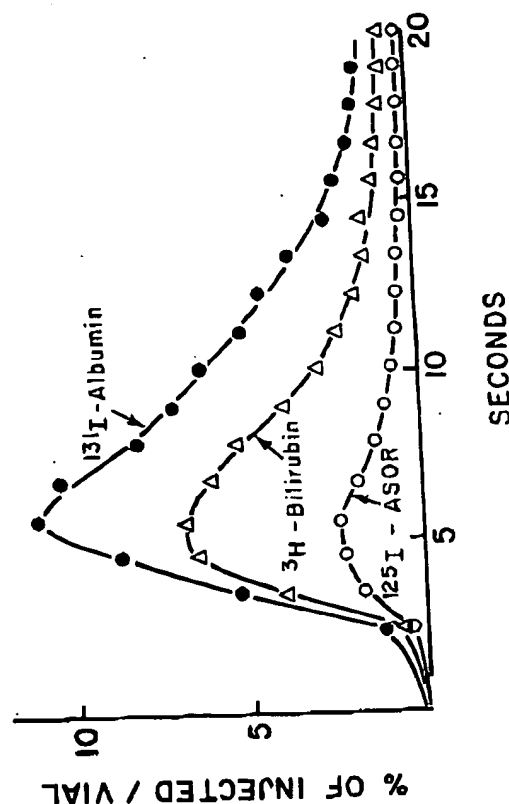


Figure 1: Hepatic venous outflow patterns of ¹³¹I-albumin, ³H-bilirubin, and ¹²⁵I-asialoorosomucoid (ASOR) following simultaneous injection into the portal vein of an isolated perfused rat liver following pre-infusion of non-immune goat IgG. (Reprinted from reference 4 with permission).

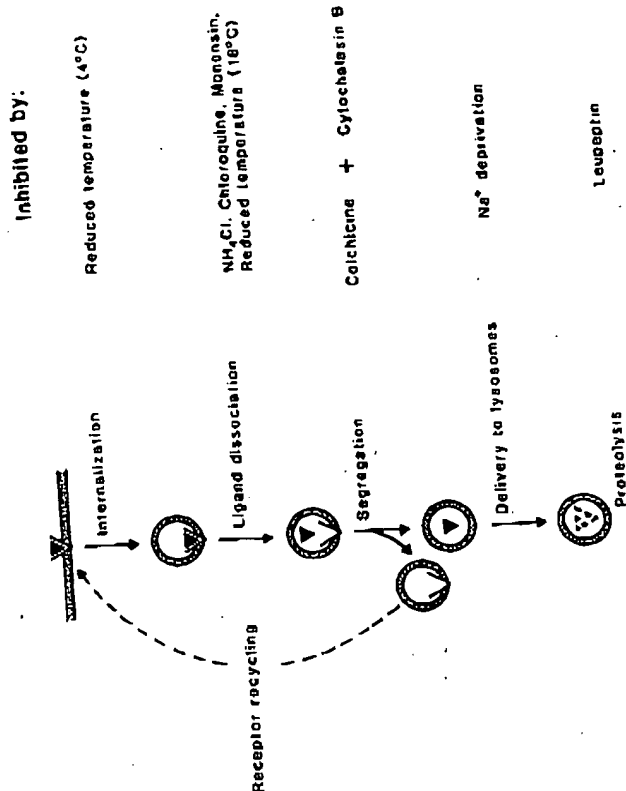


Figure 3: Schematic diagram of receptor-mediated endocytosis of ASOR and its inhibitors. Based on these and other studies, five discrete steps in uptake and catabolism can be identified. Inhibitors of each of these steps have been identified. Inhibitors are assigned on the basis of their most proximal site of action as a wave of prebound ligand moves through the pathway. (Reprinted from reference 5 with permission).

The liver cell plasma membrane plays an important role in receptor-mediated endocytosis. Because the liver cell surface may undergo marked changes during proliferation, we studied transport of ASOR and bilirubin by regenerating rat liver (7). The rat hepatocyte divides approximately once per year, and mitosis in hepatocytes is infrequently seen in normal liver (8). Following two-thirds hepatectomy, rapid cellular proliferation occurs throughout the remaining liver remnant, and is associated with expression of oncofetal antigens (9-11). Studies performed with hepatocytes in culture suggest that hepatocyte replication is associated with modulated expression of several intracellular and secreted proteins including ligandin, pyruvate kinase, and α -1-fetoprotein (12). Altered liver cell plasma membrane function during regeneration has also been suggested. Studies of the interaction of plasma membrane,

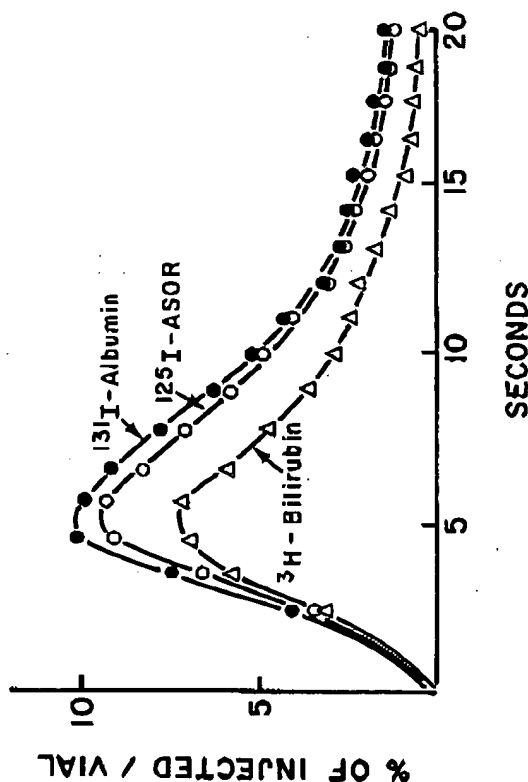


Figure 2: The same liver as in Figure 1 was then infused with anti-HBP IgG and the transport study was repeated. There was a marked reduction in uptake of ¹²⁵I-ASOR as indicated by increased recovery, while uptake of ³H-Bilirubin was unchanged. (Reprinted from reference 4 with permission).

Newer studies have revealed that endocytosis of ASOR following binding to HBP is a complex event (5). Following binding of ligand to cell surface HBP, the ligand-receptor complex is internalized into a prelysosomal compartment that has been termed the endosome. The endosome interior becomes acidified resulting in dissociation of ligand and receptor (6). The ligand and receptor segregate from each other; receptor eventually recycles to the cell surface, while ligand enters lysosomes where degradation takes place. Recent studies have identified specific inhibitors of these steps (Figure 3).

prepared from regenerating liver, with insulin and glucagon revealed an increased number of insulin receptors and reduced number of glucagon receptors (13). Amino acid uptake by hepatocytes was found to be increased several-fold during liver regeneration (14). This finding which may be due to an altered plasma membrane transport mechanism, is blocked by pretreatment with colchicine, a microtubule disrupter. Changes in other liver cell plasma membrane enzymes occur in regeneration, including a doubling of (Na^+-K^+) -ATPase activity and a reduction in glucagon-stimulated adenylyl cyclase activity (15).

As a measure of specific hepatocyte function, transport of 3H -Bilirubin and ^{125}I -ASOR was determined using the single-pass indicator dilution method in the isolated perfused regenerating liver (7). This method permits quantitation of uptake rates independent of hepatic mass. Results were compared to those obtained in sham-operated rats. As seen in Figure 4, liver weight increased progressively with time after two-thirds hepatectomy, and returned to normal by six days. Uptake of 3H -Bilirubin and ^{125}I -ASOR fell by over 50% and 80%, respectively, reaching a nadir at the time of greatest cell proliferation (Figure 5). Uptake returned to normal by six days. These studies of transport of anions and asialoglycoproteins during liver regeneration revealed functional maturation similar to that seen during development.

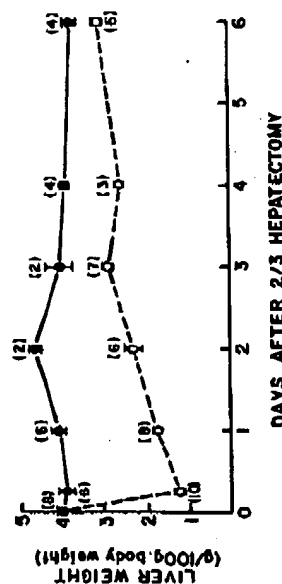


Figure 4: Liver weight in sham-operated rats (●) and two-thirds hepatectomized rats (○) at various times after surgery. (Reprinted from reference 7 with permission).

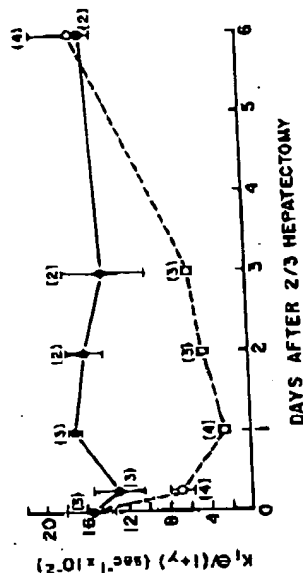


Figure 5: Influx rate of ^{125}I -ASOR ($k_1 \theta / (1 + \theta)$) in perfused liver from sham-operated (●) and partially hepatectomized rats (○). Rate constants were calculated from indicator dilution curves. Similar results were obtained in studies of 3H -Bilirubin transport. (Reprinted from reference 7 with permission).

That hepatocellular proliferation alone is not responsible for the transport alterations seen during liver regeneration was demonstrated in perfused liver from rats pretreated with nafenopin (16). Nafenopin (2-methyl-2p-(1,2,3,4-tetrahydro-1-naphthyl) phenoxy propionic acid) is a hypolipidemic drug which induces rapid liver growth characterized by hepatocellular hypertrophy and hyperplasia similar to that seen during regeneration (17-20). After nafenopin treatment, the liver has morphologic features of regeneration including proliferation of smooth endoplasmic reticulum, enlargement of peroxisomes and Golgi, and dilated and tortuous bile canaliculi (21,22). Despite a 40% increase in liver weight 24 hours after two days of nafenopin, there was no change in transport of bilirubin or ASOR, unlike results seen in regeneration (Figure 6). However, uptake of the water soluble organic anions, BSP and conjugated bilirubin was reduced by 50% (Figure 6). These studies suggest that hepatocellular proliferation alone is not responsible for the transport alterations seen during liver regeneration. Nafenopin effectively unmasks differences in uptake of bilirubin and other more water soluble organic anions such as sulfobromophthalein and conjugated bilirubin, suggesting that their uptake mechanisms are partially independent. As discussed below, reduced uptake of ASOR during liver regeneration is a consequence of reduced numbers of cell surface receptors for this ligand. Whether there are analogous alterations in organic anion interaction with liver cell surface membranes during regeneration or after nafenopin-treatment remains to be determined.

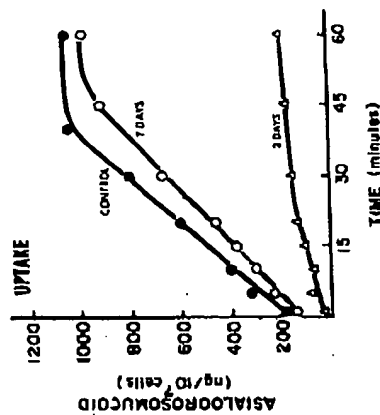


Figure 7: Uptake of ASOR by isolated hepatocytes obtained from sham-operated rats or rats 2 days (Δ) or 7 days after two-thirds hepatectomy. Similar to results in perfused liver, uptake is reduced during the proliferative phase of regeneration. (Reprinted from reference 23 with permission).

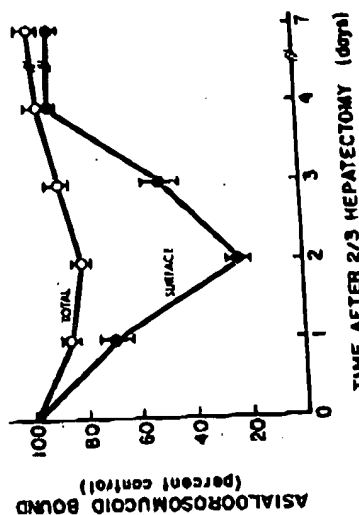


Figure 8: Binding of ASOR by intact hepatocytes (\bullet) and cell homogenates (\circ) at various times after two thirds hepatectomy. During the time of active cell proliferation, there was an 80% loss of receptor from the cell surface. (Reprinted from reference 23 with permission).

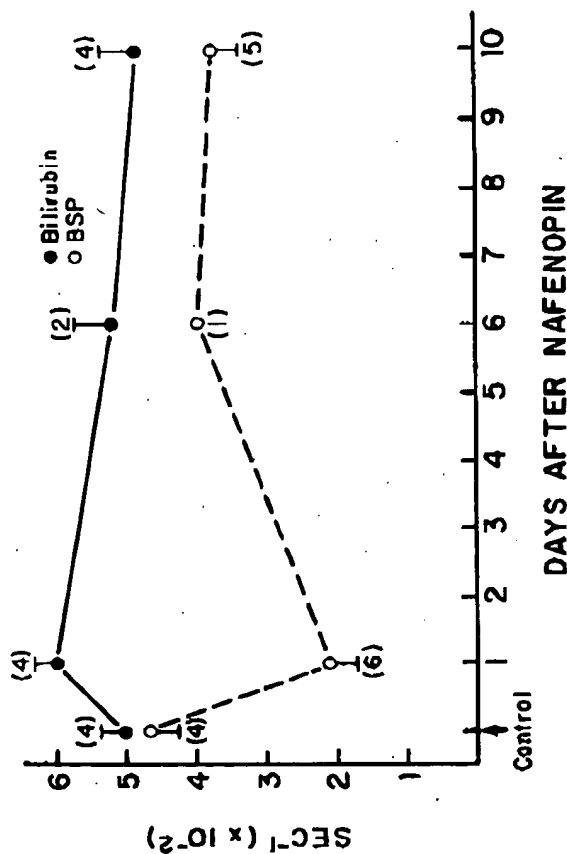


Figure 6: Influx of ³H-Bilirubin and ³⁵S-BSP in isolated perfused liver of rats pretreated with nafenopin. There was no change in influx of either compound in corn oil fed controls. Despite the marked proliferative response similar to that seen in regeneration, influx of bilirubin remained constant, as did influx of ASOR. In contrast, BSP influx was significantly reduced.

Reduced uptake of ASOR during liver regeneration could be due to a number of factors. That it is due to reduced levels of HBP on the liver cell surface, however, has been demonstrated (23). In these studies, isolated hepatocytes were prepared from livers at various times after two-thirds hepatectomy. Binding of ¹²⁵I-ASOR to the cell surface or to solubilized cell homogenates was determined as was uptake and degradation of this ligand (Figures 7 and 8). Results were compared with identical studies performed in cells obtained from sham-operated rats. Similar to results in perfused liver, there was reduced uptake of ASOR by hepatocytes obtained during the period of active cell proliferation. This was accompanied by an 80% loss of receptor from the cell surface. Total cell receptor, as determined in the solubilized homogenates, was normal (Figure 8).

These studies have suggested potential new directions in treatment of hepatocellular carcinoma. Exciting studies along these lines have recently been performed by Wu and colleagues (28) in studies of methotrexate. The lack of specificity for neoplastic tissue which results in injury to normal as well as malignant cells, has limited the clinical usefulness of this drug. In addition, hepatotoxicity frequently complicates treatment with high levels of methotrexate. These investigators synthesized a covalent conjugate of folic acid with asialofetuin with the goal of directing this methotrexate antagonist to receptor-bearing cells, sparing them from methotrexate toxicity. Less differentiated cells not containing HBP, would be killed by methotrexate.

Two cultured cell lines were used for these studies. One was a relatively undifferentiated human hepatocellular carcinoma line, PLC/PRF/5, which lacks HBP. The other was a more differentiated human hepatocellular carcinoma line, HepG2. This is the only cultured cell line which has been found to express HBP. As seen in Figure 10, PLC/PRF/5 receptor negative cells were killed by methotrexate both in the presence and absence of the asialofetuin-folic acid conjugate. Methotrexate also killed HepG2 cells, but this effect was eliminated by adding the folic acid conjugate to the medium. Thus, these studies reveal specific rescue of differentiated cells based upon the presence of a specific receptor on the cell surface. They may have important implications in the design of clinical chemotherapeutic protocols.

A similar line of investigation has been conducted on liposome delivery of drugs. Rahman and colleagues (29,30) incorporated adriamycin into liposomes composed of phosphatidylcholine and cholesterol mixed with stearyl amine (positively charged) or phosphatidylserine (negatively charged). Liposomal incorporation may result in internalization of drug into cells by endocytosis. Use of adriamycin has been limited by its cardiac toxicity. Electron microscopic studies have demonstrated degeneration of myofibrils and mitochondrial distortion, as well as a reduction in cardiac myocytes.

The modulation of liver cell HBP content seen during regeneration is similar to that which has been observed in the mouse during development (24). As seen in Figure 9, fetal mice have no detectable receptor until the nineteenth day of gestation, and develop normal adult levels by 5 days postpartum. Maternal liver has a tripling of HBP activity in the last trimester, with a fall to normal levels shortly after birth.

These studies suggested that hepatocytes during regeneration entered a state of "dedifferentiation". Other studies have revealed altered liver cell membrane enzyme activities during hepatocarcinogenesis (25). Based on these data, Stockert and Becker (26) studied HBP content of rat liver following exposure to the chemical carcinogen AAF (N-2-acetylaminofluorene). As has been described, this drug induces formation of neoplastic nodules and hepatocellular carcinoma in rat liver (27). These nodules can be dissected free of other liver tissue and studied biochemically. HBP, as assayed by specific binding of ^{125}I -ASOR, was reduced by almost 70% in neoplastic nodules and by 95% in areas of hepatocellular carcinoma.

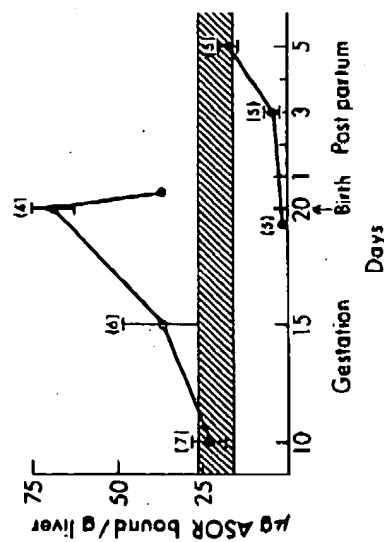


Figure 9: Asialoglycoprotein receptor binding activity in pregnancy, fetal and neonatal development. The hatched area indicates control male and virgin female mouse liver activity. Pregnant mice (○) have supranormal receptor activity while developing mice (●) do not have detectable receptor activity until the nineteenth day of gestation. (Reprinted from reference 24 with permission).

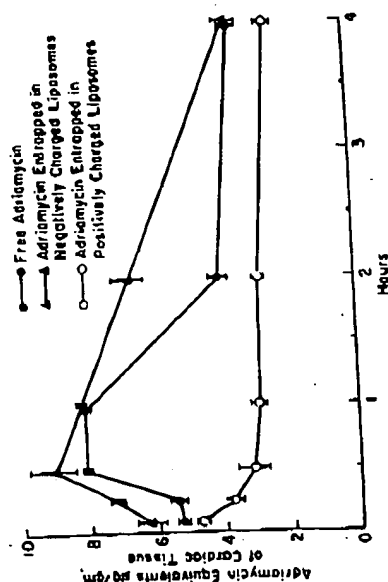


Figure 11: Adriamycin disposition in mouse heart following i.v. administration of free and liposome-entrapped drugs. (Reprinted from reference 29 with permission).

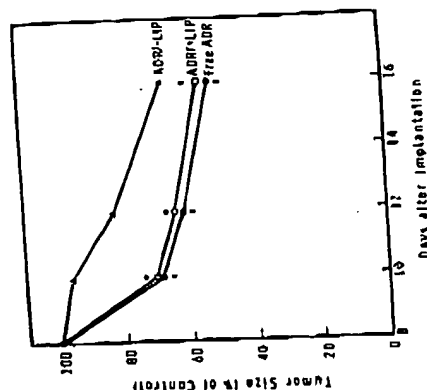


Figure 12: Treatment of mice given implants of Lewis lung carcinoma. Adriamycin (4 mg/kg) was administered i.v. to mice on days 8, 10 and 12 after tumor implantation, as free drug (Free ADR) or drug entrapped in positive (ADR/+LIP) or negative (ADR/-LIP) liposomes. The percentage of reduction of tumor mass was assessed by measuring the largest perpendicular diameter of the primary tumor. The asterisk indicates statistical difference from control ($P < 0.05$). (Reprinted from reference 29 with permission).

Pharmacokinetic studies have revealed avid uptake into heart muscle. Incorporation of adriamycin into positively charged liposomes effectively retarded the *in vivo* uptake of drug in cardiac tissue when compared to free drug or drug incorporated into negatively charged liposomes (Figure 11). In this situation, adriamycin was preferentially concentrated in liver, spleen and lungs. Electron microscopic studies revealed that the myocytes and myofibrillar structure of cardiac muscle were well preserved. Importantly, anti-tumor activity against murine ascitic P388 leukemia and Lewis lung carcinoma was identical whether adriamycin was administered alone or entrapped in positively charged liposomes (Figure 12). These studies and the studies presented above, suggest that liposomes may be developed to deliver their contents to specific cell types by targeting them to particular cell surface receptors.

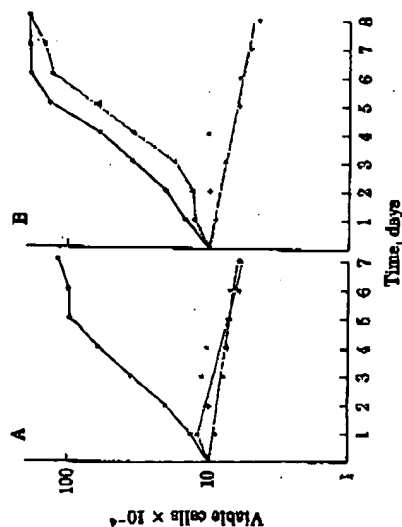


Figure 10: Specific rescue of methotrexate (MTX)-treated HBP containing cells by an asialofetuin-folic acid conjugate. (A) PLC/PRF/5 receptor-negative cells grown in the absence of MTX (○) in 0.5 μ M MTX (+), or in 0.5 μ M MTX/15 μ M asialofetuin-folic acid conjugate (Δ). (B) Hep62 receptor-positive cells grown under the same conditions. (Reprinted from reference 28 with permission).

ACKNOWLEDGEMENT: This work was supported by NIH grants AM-23026, AM-17702, AM-32419 and AM-32972.

REFERENCES

1. Stockert RJ, Morell AG. 1982. Endocytosis of Glycoproteins. In *The Liver: Biology and Pathobiology*. ed. IM Arias, H Popper, D. Schachter and DA Shafritz. Raven Press, New York, pp. 205-217.
2. Morell AG, Gregoriadis G, Scheinberg IH, Hickman J, Ashwell G. 1971. *J Biol Chem* 246:1461-1467.
3. Hudgin RL, Pricer WE Jr, Ashwell G, Stockert RJ, Morell AG. 1974. *J Biol Chem* 249:5536-5543.
4. Stockert RJ, Gartner U, Morell AG, Wolkoff AW. 1980. *J Biol Chem* 255:3830-3831.
5. Wolkoff AW, Klausner RD, Ashwell G, Harford J. 1984. *J Cell Biol* 98:375-381.
6. Harford J, Wolkoff AW, Ashwell G, Klausner RD. 1983. *J Cell Biol* 96:1824-1828.
7. Gartner U, Stockert RJ, Morell AG, Wolkoff AW. 1981. *Hepatology* 1:99-106.
8. Steiner JW, Perz ZM, Taichman LB. 1966. *Exp Mol Pathol* 5:146-181.
9. Bonney RJ, Walker PR, Potter VR. 1973. *Biochem J* 136:947-954.
10. Sell S, Nichols M, Becker FF, et al. 1974. *Cancer Res.* 34:864-871.
11. Naughton BA, Kaplan SM, Roy M, et al. 1977. *Science* 196:301-302.
12. Leffert H, Mora T, Sell S, et al. 1978. *Proc Natl Acad Sci USA* 75:1834-1838.
13. Leffert HL, Koch KS, Moran T, et al. 1979. *Gastroenterology* 76:1470-1482.
14. Walker PR, Whitefield JE. 1978. *Proc Natl Acad Sci USA* 75:1394-1398.
15. Bruscalupi G, Curatola G, Lenaz G, et al. 1980. *Biochim Biophys Acta* 597:264-273.
16. Gartner U, Stockert RJ, Levine MG, Wolkoff AW. 1982. *Gastroenterology* 83:1163-1169.
17. Hess R, Maier R, Staubli W. 1969. *Adv Exp Med Biol* 4:483-489.
18. Best MK, Duncan CH. 1970. *Atherosclerosis* 12:185-192.
19. Beckitt R, Weiss R, Stitzel R, et al. 1972. *Toxicol Appl Pharmacol* 23:43.
20. Moody DE, Rao MS, Reddy JK. 1977. *Virchows Arch B Cell Pathol* 23:291-296.
21. Novikoff AB, Novikoff PM, Mori M, et al. 1975. *J Histochem Cytochem* 23:314.
22. Leighton F, Coloma L, Koenig C. 1975. *J Cell Biol* 67:281-309.
23. Howard DJ, Stockert RJ, Morell AG. 1982. *J Biol Chem* 257:2856-2858.
24. Collins JC, Stockert RJ, Morell AG. 1984. *Hepatology* 4:80-83.
25. Gravela E, Feo F, Canuto RA, Garcea R, Gabriel L. 1975. *Cancer Res* 35:3041-3047.
26. Stockert RJ, Becker FF. 1980. *Cancer Res* 40:3632-3634.
27. Stout DL, Becker FF. 1978. *Cancer Res.* 38:2274-2278.
28. Yu GY, Wu CH, Stockert RJ. 1983. *Proc Natl Acad Sci USA* 80:3078-3080.
29. Rahman A, Kessler A, More N, Sikic B, Rowden G, Woolley P, Schein PS. 1980. *Cancer Res* 40:1532-1537.
30. Rahman A, More N, Schein PS. 1982. *Cancer Res* 42:1817-1825.